

Physico-Chemical, Phytochemical and Bioactive Characterization of *Dipteracanthus Prostratus* (Poir) Nees – An Antidote Medicinal Plant

*A. Maruthupandian¹, V. Gowthami¹, M. Kumutha¹, C. Santhosh¹ and P. Thangavel²

Received: 14 May 2015 / Accepted: 24 May 2015 / Published Online: 15 June 2015

OPEN ACCESS

©Gayathri Publishers 2015

Abstract

Dipteracanthus prostratus (Poir) Nees (Pottakanchi in tamil) belongs to Acanthaceae family and it is used as antidote for snakebite by Palliyar tribal settled in Sirumalai hills, Western Ghats, Tamil Nadu. The present investigation evaluates the physico-chemical, phytochemical, fluorescence analysis and antibacterial potential of this ethnomedicinal plant. The outcome of the study support and validate the traditional use of this plant in the treatment of various types of diseases.

Key Words: *Dipteracanthus prostratus*, Pottakanchi, Palliyar, Antibacterial, Phytochemical.

Citation: Maruthupandian, A., Gowthami, V., Kumutha, M., Santhosh, C. and Thangavel, P. 2015. Physico-Chemical, Phytochemical and Bioactive Characterization of *Dipteracanthus Prostratus* (Poir) Nees – An Antidote Medicinal Plant, 4(2):1-7.

Present Address

¹Department of Botany, School of Life Sciences, Periyar University, Periyar Palkalai Nagar, Salem – 636 011, Tamil Nadu, India.

²Department of Environmental Science, School of Life Sciences, Periyar University, Periyar Palkalai Nagar, Salem – 636 011, Tamil Nadu, India.

*Corresponding author: maruthubot@periyaruniversity.ac.in

Manuscript Type : **Manuscript**

Received Manuscript : **Via Email**

Approved Letter : **Received**

Funding Source: **Nil**

Conflict of Interest : **Nil**

Manuscript Full Responses: **Authors**



Botanical Report / © 2015 GTRP-GRF group

More information contact us botanicalreport@yahoo.com

© 2015 GTRP Reserved. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by-nd/3.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

1. Introduction

The medicinal plants are useful for healing as well as for curing of human diseases because of the presence of phytochemical constituents (Nostro *et al.*, 2000). Phytochemicals are naturally occurring in the medicinal plants, leaves, vegetables and roots that have defense mechanism and protect from various diseases. Phytochemicals are primary and secondary compounds. Chlorophyll, proteins and common sugars are included in primary constituents and secondary compounds have terpenoid, alkaloids and phenolic compounds (Krishnaiah *et al.*, 2007). These are the reservoirs of potentially useful chemical compounds which could serve as newer leads and clues for modern drug design (Vijayalakshmi and Ravindran, 2012). The most important of these bioactive constituents of plants are alkaloids, tannins, flavonoids and phenolic compounds (Doss, 2009). Correlation between the phytoconstituents and the bioactivity of plant is desirable to know for the synthesis of compounds with specific activities to treat various health ailments and chronic diseases as well (Pandey *et al.*, 2013). *Dipteracanthus prostratus* (Poir.) Nees very important traditional medicinal plant and the whole plant juice is used as a remedy for snakebite by *Palliyar* tribals in Sirumalai hills, Western Ghats, Tamil Nadu (Maruthupandian and Mohan, 2012). The aim of this study was to analysis physico-chemical, phytochemical, fluorescence and antibacterial activity of indigenous antidote medicinal plants of *Dipteracanthus prostratus* (Poir.) Nees.

2. Materials and Methods

The plant materials were collected from Sirumalai hills, Western Ghats, Tamil Nadu, India. The plant specimen was collected, air-dried and mounted on herbarium sheets and identified by using various standard floras (Fischer and Gamble, 1957; Gamble, 1957a,b; Matthew, 1983a, b, c; Nair and Henry, 1983; Henry *et al.*, 1987; Henry *et al.*, 1989; Sharma *et al.*, 1993; Sharma and Balakrishnan, 1993; Sharma and Sanjappa, 1993; Sasidharan and Sivarajan, 1996; Hajra *et al.*, 1997; Matthew, 1999a, b, c; Pallithanam, 2001), photographed and sample specimens were collected for the preparation of herbarium. The identified plant specimens were confirmed and deposited in the herbarium of Department of Botany, School of Life Sciences, Periyar University, Salem, Tamil Nadu.

2.1 Physico-chemical and Fluorescence analysis

These studies were carried out as per the standard procedures (Lala, 1993). In the present study, the powdered whole plant was treated with various chemical reagents like aqueous 1N sodium hydroxide, alcoholic 1N sodium hydroxide, 1N hydrochloric acid, 50% sulphuric acid, concentrated nitric acid, picric acid, acetic acid, ferric chloride and concentrated HNO_3+NH_3 . These extracts were subjected to fluorescence analysis in day light and UV light (254nm and 366nm). Various ash types and extractive values were determined by standard methods (Anonymous, 1996).

2.2 Phytochemical analysis

Shade dried and powdered samples were successively extracted with chloroform, ethanol and petroleum ether. The extracts were filtered ad concentrated using vacuum distillation. The different extracts were subjected to qualitative tests for the identification of various phytochemical constituents as per the standard procedure (Lala, 1993 and Brindha *et al.*, 1981).

2.3 Antibacterial Assay

2.3.1 Collection of microorganisms

Stock cultures of bacteria such as *Staphylococcus aureus*, *Bacillus subtilis*, *Pseudomonas aeruginosa* and *Escherichia coli* were obtained from Research Laboratory, Department of Microbiology, School of Biosciences, Periyar University, Salem, Tamil Nadu.

2.3.2 Preparation of media

The growth media employed in the present study included Nutrient agar and Nutrient broth. The medium was adjusted to pH 7.4 and sterilized by autoclaving at 15 lbs pressure (121°C) for 15 minutes.

2.3.3 Sub culturing of microorganisms

The pure culture of microorganism was maintained on nutrient agar slants by frequent sub culturing. The culture was stored at 4°C.

2.3.4 Preparation of inoculum

Each organism was recovered for testing by sub culturing on fresh media. A loopful inoculum of each bacterium was suspended in 5ml of nutrient broth and incubated overnight at 37°C. These overnight cultures were used as inoculum.

2.3.5 Antibacterial activity

Antibacterial activity was demonstrated by modification of the method described by Barry and Thornsberry, (1985). 0.1 ml of the diluted microbial culture was spread on sterile nutrient agar plate. The pre-soaked and dried discs of 6mm diameter of What man No.1 filter paper were then placed on the seeded plates and gently pressed down to ensure contact. At the same time standard antibiotic of Tetracycline (30 μ g/ disc) was used as reference or positive control. Respective solvents without plant extracts served as negative control. The plates were incubated at 37°C for 24 hours. After the incubation period, the diameter of the inhibition zone around the plant extract saturated discs were measured and also compared with the diameter of inhibition zone of commercial standard antibiotic discs. The inhibition zone around the discs were measured and recorded as the difference in diameter between the disc (6mm) and growth free zone.

3. Results And Discussion

3.1 Physicochemical constants (Ash and Extractive values)

The result of the ash and extractive values of whole plant of *Dipteracanthus prostratus* are represents in table 1.

Table 1: Ash and extractive values of powdered whole plant of *Dipteracanthus prostratus*

Sl. No.	Type of ash	Ash value (%)
1	Total ash	6.78 \pm 0.12
2	Water soluble ash	2.19 \pm 0.05
3	Acid insoluble ash	2.41 \pm 0.37
4	Sulphated ash	11.13 \pm 0.22

Table -2 : Fluorescence analysis of the powdered in whole plant of *Dipteracanthus prostratus*

Sl.No.	Experiments	Visible / Day light	UV - light	
			254nm (short wave length)	365nm (long wave length)
1	Drug powder as such	Green	Dark green	Light green
2	Powder +1N NaOH (Aqueous)	Dark green	Green	Light green
3	Powder +1N NaOH (Alcohol)	Dark green	Green	Light green
4	Powder +1N HCL	Dark green	Green	Fluorescence green
5	Powder +50% H ₂ SO ₄	Dark green	Fluorescence green	Light green
6	Powder + Nitric acid	Red	Red	Light brown
7	Powder +Acetic acid	Green	Dark green	Pink
8	Powder +5% Ferric chloride	Dark green	Dark green	Dark green
9	Powder + HNO ₃ + NH ₃	Light green	Green	Light green

Table 3: Preliminary phytochemical screening of whole plant of *Dipteracanthus prostratus*

Pytochemical Test	Petroleum Ether	Chloroform	Ethanol
Alkaloid	+	+	+
Anthraquinone	-	-	-
Coumarin	+	-	-
Flavonoid	-	-	+
Phenol	-	-	+
Protein	-	-	-
Saponin	-	-	+
Steriod	+	+	-
Sugar	+	+	+
Tannin	+	+	+

Many phytocompounds fluoresce when suitably illuminated. The fluorescence colour is specific for each compound. A non fluorescent compound may fluoresce if mixed with impurities that are fluorescent. The fluorescent method is adequately sensitive and enables the precise and accurate determination of the analyze over a satisfactory concentration range without several time consuming dilution steps prior to analysis of pharmaceutical samples (Pimenta *et al.*, 2006).

3.3 Preliminary phytochemical analysis

The chloroform extract of the whole plant of *Dipteracanthus prostratus* shows the presence of alkaloid, steroid, sugar and tannins. The petroleum ether extracts of whole plant of *Dipteracanthus prostratus* shows alkaloid, coumarin, steroid, sugar and tannin. The ethanol extracts of whole plant of *Dipteracanthus prostratus* depicts alkaloid, flavonoid, phenol, saponin, steroid, sugar and tannin.

Presence or absence of certain important compounds in an extract is determined by colour reactions of the compounds with specific chemicals which act as dyes. This procedure is a simple preliminary prerequisite before going for detailed phytochemical investigation. Various tests have been conducted qualitatively to find out the presence or absence of bioactive compounds. Different chemical compounds such as alkaloids, coumarin, flavonoid, phenol, protein, saponin, steroid, sugar and tannin were detected in *Dipteracanthus prostratus* extracts which could make the plant useful for treating different ailments as having a potential of providing useful drugs of human use. This is because the pharmacological activity of any plant is usually traced to a particular compound.

To understand the nature of the fluorescence emission from these crude preparations under different conditions, the preliminary phytochemical analysis of

these crude preparations was compared. The comparative analysis clearly showed a correlation between a compound present in it and their fluorescent behavior under different conditions. The major bioactive compounds present in these crude preparations are the coumarin, flavones, tannins, alkaloids and saponin. Coumarin especially hydroxyl amino acid derivatives like o-coumaric acid appears yellowish green in alkaline condition under short UV radiation. Flavones which are light yellow in aqueous condition under UV light turns to bright yellow under alkaline conditions. Similarly the phytosterols when treated with 50% H₂SO₄ show green fluorescence under UV light. Terpenoids especially sapogenins exhibit yellow green fluorescence under short UV light (Horborne, 1976). Quinine, aconitin, berberin and emetin show specific colour of fluorescence (Aconitin - light blue; berberin - light yellow; emetin - orange). Fixed oils and fats fluoresce least, waxes more strongly and mineral salts most of all (Evans, 1996). Haydon (1975) studied the photophysical characters of coumarins. Hydroxy methyl coumarin fluoresced in the 420 - 440nm when observed in different solvents with increasing polarity (Chaltpudhyay, 2006). The fluorescence analysis of the crude drugs of *Dipteracanthus prostratus* exhibited clear fluorescence behaviour at different radiations which can be taken as standard fluorescence pattern. The physic-chemical constant evaluation of the drug is an important parameter in detecting adulteration or improper handling of drugs. The phytochemicals revealed that these plants contain bioactive substances such as alkaloids, coumarin, flavonoid, saponin, steroids, phenols, sugars and tannins. Flavonoids and phenols have been reported to exert multiple biological effects such as anti-inflammatory, antiallergic, antioxidant, antidiabetic and cancer activities. The curative properties of the medicinal plants are due to the presence of various secondary metabolites.

3.5 Antibacterial activity

The present investigation, the antibacterial activity evaluated chloroform, ethanol and petroleum ether extracts of *Dipteracanthus prostratus* against two gram positive bacteria (*Staphylococcus aureus* and *Bacillus subtilis*) and two gram negative bacteria (*Pseudomonas aeruginosa* and *Escherichia coli*) used with antibiotic tetracycline. Chloroform extracts of whole plant of *Dipteracanthus prostratus* showed maximum inhibitory activity was observed against *Staphylococcus aureus* (4mm). Moderate activity was observed against *Bacillus subtilis* (3.6mm) and *Staphylococcus aureus* (3mm). Ethanol extracts of whole plant powder of *Dipteracanthus prostratus* showed maximum inhibitory activity was observed against *Staphylococcus aureus* (5.2mm). Moderate activity was observed against *Bacillus subtilis* (5.1mm). Petroleum ether extracts of whole plant powder of *Dipteracanthus prostratus* showed maximum inhibitory activity was observed against *Staphylococcus aureus* (5mm) moderate activity observed against *Bacillus subtilis* and *Pseudomonas aeruginosa* (4.6mm).

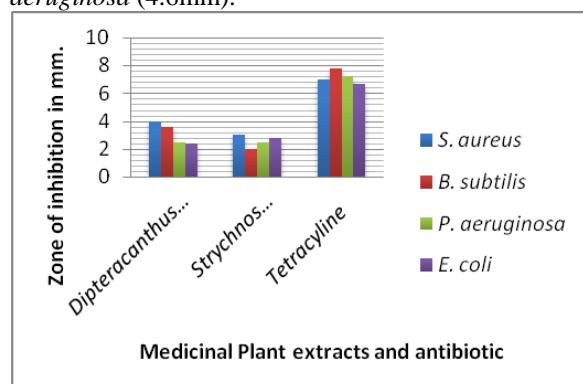


Fig. 1: Antibacterial activity of chloroform extract of *Dipteracanthus prostratus*

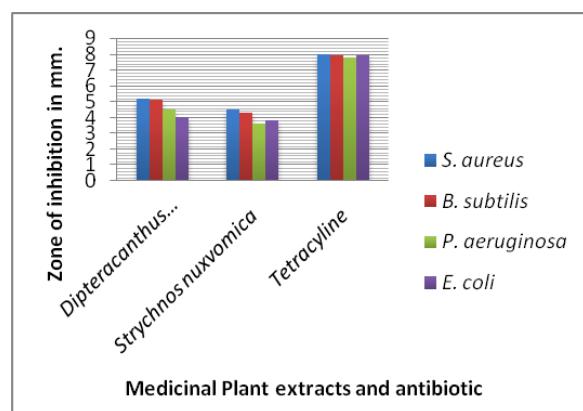


Fig. 2: Antibacterial activity of ethanol extract of *Dipteracanthus prostratus*

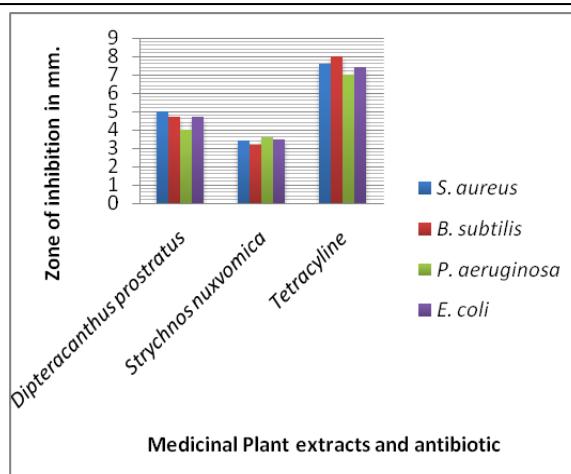


Fig.3: Antibacterial activity of Petroleum ether extract of *Dipteracanthus prostratus*

Among the bacterial organisms tested, the chloroform, petroleum ether and ethanol extracts of whole plant of *Dipteracanthus prostratus* exhibit maximum activity with showed higher inhibitory activity with *Staphylococcus aureus*, a gram positive bacterium. The three extracts of above said plants inhibited the growth of this organisms compared with tetracycline as positive control, a standard antibiotic used. The results obtained are encouraging as the chloroform and ethanol extracts have shown considerable antibacterial activity against tested organisms. The antibacterial activities of the plants may be attributed to the presence of bioactive principles such as phenols, steroids, alkaloids and flavonoid as suggested by several literatures.

3.6 Conclusion

The results were recommend in this plant of *Dipteracanthus prostratus* (Poir) Nees can be act as a potential source of bioactive compounds which could be used as effective drug to cure various diseases after completion of further pharmacological studies and prepare active principles with drug standardization.

4. References

- African Pharmacopoeia. 1986. General Methods for Analysis Is ted. 2: (OAU/STRC) Lagos. P. 123.
- Anonymous, 1996. Indian Pharmacopoea, Vol. I & II. Government of India, Ministry of Health.
- Barry A.L. and Thornsberry, C.1985. Susceptibility tests. Disc diffusion test procedures. In: Manual of Clinical Microbiology (Eds). Lennette Balows, EH.,

W.J. Hauster and H.J. Shadomy. American Society for Microbiology, Washington, DC. (1985) 978-987.

Brindha, P., Sasikala, P. and Purushothaman, K.K. 1981. Pharmacognostic studies on Merugan Kizhangu. *Bull. Med. Eth. Bot. Res.*, 3: 84-96.

Chaltopudhyay, N., Mallick, A. and Sengupta, S. 2006. Photophysical studies of hydroxyl methyl coumarin: A new fluorescent chemosensor for Zinc and Nickel ions in water. *J. Photochem. Photobiol.*, 177: 55-60.

Evans, W.C. 1996. In: Trease and Evans' Pharmacognosy. Singapore, Harcourt Baraco and Company Asia Pvt. Ltd. pp. 1-437.

Fischer C.E.C. and Gamble J.S. 1957. The Flora of the Presidency of Madras III: (Rep. Ed.): 1347-2017.

Gamble, J.S. 1957a. The Flora of the Presidency of Madras I: (Rep. Ed.): 1-577.

Gamble, J.S. 1957b. The Flora of the Presidency of Madras II: (Rep. Ed.) : 578-1346.

Ganesan S. and Suresh, N. and Kesavan L. 2004. Ethnomedicinal survey of lower Palni Hills of Tamil Nadu. *Ind. J. Trad. Knowled.*, 3: 299-304.

Hajra, P.K., Nair, V.J. and Daniel P. 1997. Flora of India. IV: 1 – 561.

Haydon, S.C. 1975. *Spectroscopic letters*, 8: 815

Henry, A.N., Chithra V. and Balakrishnan N.P. 1989. Flora of Tamil Nadu, India, Series I: Analysis III: 1-171.

Henry, A.N., Kumari, G.R. and Chithra, V. 1987. Flora of Tamil Nadu, India, Series I: Analysis II: 1-258.

Horborne, J.B. 1976. Phytochemical methods. Chapman & Hall, New York. pp. 1- 288.

Krishnaiah, D., Sarbatly, R. and Bono, A. 2007. Phytochemical antioxidants for health and medicine: A move towards nature. *Biotechnol. Mol. Biol. Rev.*, 1: 97-104.

Lala, P.K. 1993. Lab Manuals of Pharmacognosy. CSI Publishers and distributors, Calcutta, 5th Edition. 1993.

Maruthupandian, A, Mohan. V. R. and Kottaimuthu, 2011. Ethnomedicinal plants used for the treatment of diabetes and jaundice by palliyar tribals in sirumalai hills, Western Ghats, Tamil Nadu, India. *Ind. J. Nat. Prod. Resourc.*, 2: 493-497.

Matthew K.M. (983a. The flora of the Tamil Nadu Carnatic I: 1- 688.

Matthew K.M. 1983b. The flora of the Tamil Nadu Carnatic II: 689- 1540.

Matthew K.M. 1983c. The flora of the Tamil Nadu Carnatic III: 1541- 2155.

Matthew K.M. 1999a. The flora of the Palni Hills I: 1- 575.

Matthew K.M. 1999b. The flora of the Palni Hills II: 576- 1196.

Matthew K.M. 1999c. The flora of the Palni Hills III: 1197- 1635.

Musa, K.Y., Katsayal, A.U., Ahmed, A., Mohammed, Z. and Danmalam., U. H. 2006. Pharmacognostic investigation of the leaves of *Gisekia Pharmacoides*. *African J. Biotech.*, 5: 956-957.

Nair N.C. and Henry A.N. 1983. Flora of Tamil Nadu, India, Series I: Analysis I: 1-184.

Nostro, A., Germanò, M.P., D'angelo, V., Marino, A. and Cannatelli, M.A. 2000. Extraction methods and bioautography for evaluation of medicinal plant antimicrobial activity. *Lett. Appl. Microbiol.*, 30: 379-384.

Pallithanam, J.M. 2001. A Pocket flora of the Sirumalai hills, South India: i-360.

Pandey, P., Mehta, R. and Upadhyay, R. 2013. Physico-chemical and preliminary phytochemical screening of *Psoralea corylifolia*. *Arch. Appl. Sci. Res.*, 5:261-265.

Pimenta, A. M., Montenegro, M.C, Ara Ujo A.N. and Mart'inez, J.C. 2006. Application of sequential injections analysis to Pharmaceutical analysis. *J. Pharm. Biomed. Anul.*, 40: 16-34

Pushpangadan P. and Atal, C.K. 1984. Ethno-medico-botanical investigations in Kerala I. Some primitive tribal of Western Ghats and their herbal medicine. *J. Ethnopharmacol.*, 11: 59-77.

Sharma, B.D. and Balakrishnan, N.P. 1993. Flora of India II: 1-625.

Sharma, B.D. and Sanjappa, M. 1993. Flora of India III: 1-639.

Sharma, B.D., Balakrishnan, N.P., Rao, R.R. and Hajra P.K. 1993. Flora of India I: 1- 467.

Vijyalakshmi, R., Ravindran, R. 2012. Preliminary comparative phytochemical screening of root extracts of *Diospyrus ferrea* (Wild.) Bakh and *Arva lanata* (L.) Juss. Ex Schultes. *Asian J. Plant Sci. Res.*, 2:581-587.

Doss, A. 2009. Preliminary phytochemical screening of some Indian medicinal plants. *Anc. Sci. Life*, 29:12-16.